



SYSTEMATIC REVIEW PROTOCOL FOR ANIMAL INTERVENTION STUDIES

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VERSION 2.0 (DECEMBER 2014)

| Item # | Section/Subsection/Item | Description | Check for approval |
|----------------------|---|---|--------------------|
| A. General | | | |
| 1. | Title of the review | Development of immunity in animals following Anti-rabies vaccination: a systematic review and meta-analysis | |
| 2. | Authors (names, affiliations, contributions) | Hasanthi Rathnadiwakara ¹ , Chandrindu Abeykoon ² , Ranil Jayawardena ¹ , Mangala Gunatilake ¹ ¹ Department of Physiology, Faculty of Medicine, University of Colombo, Sri Lanka ² Department of Veterinary Clinical Science, University of Peradeniya | |
| 3. | Other contributors (names, affiliations, contributions) | Merel Ritskes- Hoitinga (Professor, SYRCLE team) Judith van Luijk (Postdoc, SYRCLE team) | |
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| 5. | Funding sources/sponsors | None | |
| 6. | Conflicts of interest | None | |
| 7. | Date and location of protocol registration | June 2020 www.syrcle.nl | |
| 8. | Registration number (if applicable) | | |
| 9. | Stage of review at time of registration | | |
| B. Objectives | | | |
| Background | | | |
| 10. | What is already known about this disease/model/intervention? Why is it important to do this review? | Vaccination of animal species responsible for transmission of rabies virus has become the most effective method to control rabies. Studies have shown that anti-rabies vaccination induce humoral and cellular immune response. However, the efficacy of the vaccine and subsequent immune response can be influenced by various factors. Assessment of this efficacy is largely measured in animals by determination of virus neutralising antibody titres which indicates humoral immunity. Initially rabies-virus neutralising antibodies had been measured in vivo using the mouse neutralization test. Currently there are several methods available for the determination of humoral immunity such as rapid fluorescent focus inhibition test (RFFIT), Fluorescent antibody virus neutralisation (FAVN) test and semi quantitative ELISA methods. Regarding cellular immunity detection, the techniques such as ELISpot assay, Luminex technique and MTT colorimetric assays are being used. | |

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| | | Since there are different immunity detection methods used in different countries, we are planning to compare all these methods with respect to the country, type/brand of vaccine used, type of animal model tested. We are planning to assess the reliability of each method and the presence of drawbacks. Also, we are planning to find whether there are any factors that affect the efficacy and immunity development following vaccination (such as animal dependant factors). | |
| Research question | | | |
| 11. | Specify the disease/health problem of interest | Rabies | |
| 12. | Specify the population/species studied | Mammals | |
| 13. | Specify the intervention/exposure | Anti-rabies vaccination | |
| 14. | Specify the control population | Control populations as indicated in the studies e.g. un-vaccinated control groups and ect | |
| 15. | Specify the outcome measures | Humoral and cellular immunity development | |
| 16. | State your research question (based on items 11-15) | What is the status of humoral and cellular immunity development in animals against rabies following anti-rabies vaccination? | |
| C. Methods | | | |
| Search and study identification | | | |
| 17. | Identify literature databases to search (e.g. Pubmed, Embase, Web of science) | <input checked="" type="checkbox"/> PubMed <input checked="" type="checkbox"/> Web of Science <input checked="" type="checkbox"/> SCOPUS <input type="checkbox"/> EMBASE <input type="checkbox"/> Other, namely: Non bibliographical database (Google) <input type="checkbox"/> Specific journal(s), namely: | |
| 18. | Define electronic search strategies | Please see below the protocol | |
| 19. | Identify other sources for study identification | <input checked="" type="checkbox"/> Reference lists of included studies <input type="checkbox"/> Books <input type="checkbox"/> Reference lists of relevant reviews <input checked="" type="checkbox"/> Conference proceedings, namely: Sri Lanka Veterinary Association <input type="checkbox"/> Contacting authors/ organisations, namely: <input type="checkbox"/> Other, namely: | |
| 20. | Define search strategy for these other sources | Screening the reference list for relevant titles and screening the abstract of those relevant titles | |
| Study selection | | | |
| 21. | Define screening phases (e.g. pre-screening based on title/abstract, full text screening, both) | 1. Pre-screening based on title and abstract 2. Full text screening of relevant articles selected from first phase | |
| 22. | Specify (a) the number of reviewers per screening phase and (b) how discrepancies will be resolved | (a) Two reviewers will independently screen title and full text for the inclusion criteria (b) Differences in opinion that cannot be resolved by discussion will be resolved by involving the third investigator | |
| <i>Define all inclusion and exclusion criteria based on:</i> | | | |

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| 23. | Type of study (design) | Inclusion criteria: Animal intervention (studies will be included regardless of the methodology and quality) Exclusion criteria: Non-intervention studies | |
| 24. | Type of animals/population (e.g. age, gender, disease model) | Inclusion criteria: All animal models for rabies (Regardless of age, gender, species, housing/setting) Exclusion criteria: Human, Animals with pathological conditions | |
| 25. | Type of intervention (e.g. dosage, timing, frequency) | Inclusion criteria: Anti-rabies vaccination (All routes of administration, frequency, types, doses and combine vaccines) Exclusion criteria: Natural infection and other type of medical intervention (i.e: Other treatments/ drug administrations) | |
| 26. | Outcome measures | Inclusion criteria: Humoral immunity; Antibody titres (IU/ml) in FAVN, RFFIT. Presence or absence of antibodies in ELISA, detection of antibodies/ humoral immune response by any other accepted method Cellular immunity; IFN γ secreting cell count, Plasma cells and memory B cell count, Cytokine profiles in peripheral blood mononuclear cells (pg/ml or any other relevant unit) Exclusion criteria: Cellular immunity detection alone or other outcomes such as clinical signs | |
| 27. | Language restrictions | Inclusion criteria: English. If studies found in other languages, those will be translated to English using Google translator and the necessary information will be gathered Exclusion criteria: | |
| 28. | Publication date restrictions | Inclusion criteria: All publication dates Exclusion criteria: None | |
| 29. | Other | Inclusion criteria: Primary studies, Studies that present original work Exclusion criteria: Non-primary studies, duplicate studies | |
| 30. | Sort and prioritize your exclusion criteria per selection phase | Selection phase tiab screening: 1. Not an animal study (Human study) 2. Not a primary study (review articles) 3. Not an intervention study (not given the rabies vaccine) Selection phase: 1. Other type of intervention, apart from vaccination 2. Different outcome measure (other than immunity) 3. Presence of pathological conditions in the study group 4. Duplicate study/ repeated studies in the same conditions, same country | |
| Study characteristics to be extracted (for assessment of external validity, reporting quality) | | | |
| 31. | Study ID (e.g. authors, year) | Authors, year, country, journal | |
| 32. | Study design characteristics (e.g. experimental groups, number of animals) | Animal model tested, Number of animals used | |
| 33. | Animal model characteristics (e.g. species, gender, disease induction) | Species, age, gender, random stray population/ domesticated population/ laboratory model, health status | |

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| 34. | Intervention characteristics (<i>e.g.</i> intervention, timing, duration) | Type of vaccine given (combined vaccines/ different brands), number of booster doses given, age of vaccination, route of administration, | |
| 35. | Outcome measures | Frequency of sample collection, time separation between vaccination and 1 st measurement, immunity level at each time points, method used to detect immunity, sample specifications (any criteria that was used to select samples for testing; eg; If haemolysed samples were tested or not) | |
| 36. | Other (<i>e.g.</i> drop-outs) | Any confounding factors to immunogenicity, important finding/ conclusion, future directions | |
| Assessment risk of bias (internal validity) or study quality | | | |
| 37. | Specify (a) the number of reviewers assessing the risk of bias/study quality in each study and (b) how discrepancies will be resolved | (a) Two reviewers will independently assess the bias (b) Differences that cannot be resolved by discussion will be resolved by involving the third investigator | |
| 38. | Define criteria to assess (a) the internal validity of included studies (<i>e.g.</i> selection, performance, detection and attrition bias) and/or (b) other study quality measures (<i>e.g.</i> reporting quality, power) | <input checked="" type="checkbox"/> By use of SYRCLE's Risk of Bias tool <input type="checkbox"/> By use of SYRCLE's Risk of Bias tool, adapted as follows: <input type="checkbox"/> By use of CAMARADES' study quality checklist, e.g.²² <input type="checkbox"/> By use of CAMARADES' study quality checklist, adapted as follows: <input type="checkbox"/> Other criteria, namely: | |
| Collection of outcome data | | | |
| 39. | For each outcome measure, define the type of data to be extracted (<i>e.g.</i> continuous/dichotomous, unit of measurement) | <ul style="list-style-type: none"> • Frequency of sample collection; number of samples collected after vaccination in each time interval-continuous data • Time separation between vaccination and 1st measurement; number of days – continuous data • Immunity level at each time points: Humoral immunity by antibody titre (IU/ml) – continuous data Presence or absence of antibodies in ELISA (dichotomous) and their respective percentages (continuous) Cellular immunity by different mononuclear cell counts and cytokine profiles (continuous data in pg/ml or any other relevant unit) | |
| 40. | Methods for data extraction/retrieval (<i>e.g.</i> first extraction from graphs using a digital screen ruler, then contacting authors) | Extract data from tables, graphs (simple screen ruler) Contacting authors in case of missing data (only if the author contact details are available) | |
| 41. | Specify (a) the number of reviewers extracting data and (b) how discrepancies will be resolved | (a) Two authors will extract data independently (b) Differences will be resolved by discussion | |
| Data analysis/synthesis | | | |
| 42. | Specify (per outcome measure) how you are planning to combine/compare the data (<i>e.g.</i> descriptive summary, meta-analysis) | Meta-analysis with subgroup analysis for both outcome measures | |

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| 43. | Specify (per outcome measure) how it will be decided whether a meta-analysis will be performed | If the studies are sufficiently comparable (based on the design etc), outcome data will be pooled. Minimum 3 studies will be considered If not comparable, subgroup analysis will be formed | |
| <i>If a meta-analysis seems feasible/sensible, specify (for each outcome measure):</i> | | | |
| 44. | The effect measure to be used (e.g. mean difference, standardized mean difference, risk ratio, odds ratio) | Humoral immunity measurement: Standardized mean difference of antibody titres. All data converted to (IU/ml) Cellular immunity: Standardized mean difference of number of cells and different cytokine concentrations (depending on the reported units whether they can be converted into a single unit) | |
| 45. | The statistical model of analysis (e.g. random or fixed effects model) | Random effects model | |
| 46. | The statistical methods to assess heterogeneity (e.g. I^2 , Q) | I^2 | |
| 47. | Which study characteristics will be examined as potential source of heterogeneity (subgroup analysis) | Species variation Gender variation Type of vaccine (e.g. inactivated, killed etc)/ route of administration Duration of immunity development Different brands of anti-rabies vaccines used (The analysis will be done with respect to different manufacturers, and details will be collected about the sponsors and collaborators of those studies as well) | |
| 48. | Any sensitivity analyses you propose to perform | During the review process, if any individual variations/ specialities of the studies which can influence the finding of the review are identified, a suitable sensitivity analysis will be performed (eg: differences in the number of booster vaccinations used, use of a different kind of animal model/ species which do not used commonly etc) | |
| 49. | Other details meta-analysis (e.g. correction for multiple testing, correction for multiple use of control group) | If multiple testing available, we will make subgroups with similar testing and analysis will be done. Similarly, subgroup analysis will be done based on specific control groups used in the studies. | |
| 50. | The method for assessment of publication bias | Funnel plot will be drawn | |
| Final approval by (names, affiliations): Date: | | | |

Search Strings

| Database | Search equation | Number of hits (14/06/2020) |
|----------|--|--------------------------------|
| PubMed | ((rabies[Title/Abstract]) AND (immunity[Title/Abstract])) AND ((vaccine[Title/Abstract]) OR (vaccination[Title/Abstract])) | 416 |

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|----------------|---|------|
| Web of Science | #1. [vaccine AND vaccination] #2. [rabies AND immunity] #3. #2 AND #1 | 313 |
| Scopus | (((TITLE-ABS-KEY (rabies) AND TITLE-ABS-KEY (immunity)) AND ((TITLE-ABS-KEY (vaccine) OR TITLE-ABS-KEY (vaccination)))) | 1124 |